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December 14, 2001

BOX PCT

Commissioner for Patents
Washington, D.C. 20231

PCT/JP00/03896
-filed June 15, 2000

Re: Application of Shingo KATO, Yoshio KOBAYASHI, Yoshiyuki HIRAISHI, Kayoko SHIMIZU and Tetsuyoshi SUGITA
METHOD FOR DETERMINING HIV-1 SUBTYPE
Assignee: OTSUKA PHARMACEUTICAL CO., LTD. and
KEIO UNIVERSITY
Our Ref: Q67685

Dear Sir:

The following documents and fees are submitted herewith in connection with the above application for the purpose of entering the National stage under 35 U.S.C. § 371 and in accordance with Chapter II of the Patent Cooperation Treaty:

- ☒ an executed Declaration and Power of Attorney.
- ☒ an English translation of the International Application.
- ☒ ten (10) sheets of drawings.
- ☒ twelve (12) pages of Sequence Listing and a 3.5" disk containing Sequence Listing.
- ☒ Statement in Support of Submission in Accordance with 37 C.F.R. § 1.821.
- ☒ an executed Assignment and PTO 1595 form.
- ☒ Information Disclosure Statement and a Form PTO-1449.

It is assumed that copies of the International Application, the International Search Report, the International Preliminary Examination Report, and any Articles 19 and 34 amendments as required by § 371(c) will be supplied directly by the International Bureau, but if further copies are needed, the undersigned can easily provide them upon request.

The Government filing fee is calculated as follows:

Total claims	<u>18</u>	-	<u>20</u>	=	<u> </u>	x	\$18.00	=	<u>\$0.00</u>
Independent claims	<u>2</u>	-	<u>3</u>	=	<u> </u>	x	\$84.00	=	<u>\$0.00</u>
Base Fee									<u>\$890.00</u>

TOTAL FILING FEE
Recordation of Assignment
TOTAL FEE

<u>\$890.00</u>
<u>\$ 40.00</u>
<u>\$930.00</u>



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Checks for the statutory filing fee of \$890.00 and Assignment recordation fee of \$40.00 are attached. You are also directed and authorized to charge or credit any difference or overpayment to Deposit Account No. 19-4880. The Commissioner is hereby authorized to charge any fees under 37 C.F.R. §§ 1.16, 1.17 and 1.492 which may be required during the entire pendency of the application to Deposit Account No. 19-4880. A duplicate copy of this transmittal letter is attached.

Priority is claimed from:

<u>Country</u>	<u>Application No</u>	<u>Filing Date</u>
Japan	1999-167736	June 15, 1999
Japan	2000-23581	February 1, 2000

Respectfully submitted,



Gordon Kit

Registration No. 30,764

GK/amt

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

Shingo KATO et al

CHAPTER II of
Appln. No.: PCT/JP00/03896

Group Art Unit: 0000

Filed: December 14, 2001

Examiner: Unknown

For: METHOD FOR DETERMINING HIV-1 SUBTYPE

**STATEMENT IN SUPPORT OF SUBMISSION
IN ACCORDANCE WITH 37 C.F.R. § 1.821**

Assistant Commissioner
of Patents
Washington, D.C. 20231

Sir:

The following statement is provided to meet the requirements
of 37 C.F.R. § 1.821.

I hereby state that the content of the computer readable copy
(PatentIn Version 2.1) of the Sequence Listing (attached hereto)
submitted in accordance with 37 C.F.R. §§ 1.821(c) and (e),
respectively, is the same as the Sequence Listing submitted
herewith.

I hereby declare that all statements made herein of my own
knowledge are true and that all statements made on information and
belief are believed to be true; and further that these statements
were made with the knowledge and that willful false statements and
the like so made are punishable by fine or imprisonment, or both,
under Section 1001 of Title 18 of the United States Code, and that
such willful false statements may jeopardize the validity of this
application or any patent issuing thereon.

12/14/01

Date


Gordon Kit

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DESCRIPTION

Method for Determining HIV-1 SubtypeTECHNICAL FIELD

The present invention relates to a method for
5 determining HIV-1 subtypes, and a kit for determining HIV-
1 subtypes.

BACKGROUND ART

The virus causing acquired immune deficiency
syndrome (AIDS) is the human immunodeficiency virus (HIV),
10 of which type 1 (HIV-1) and type 2 (HIV-2) are known.
Most cases involve HIV-1, for which various subtypes have
been discovered.

Determining the HIV-1 subtype in infected
individuals is important for assessing the route of
15 infection and the reliability of virological test results
(particularly the drug resistance based on genotype or the
determination of plasma HIV-1 RNA concentration). HIV-1
subtypes are generally determined through the sequencing
of specific regions of the virus genome and phylogenetic
20 analysis of the results, but these are complicated and
expensive procedures.

An object of the present invention is thus to
provide a simpler method for determining HIV-1 subtypes.

Another object of the present invention is to
25 provide a kit for determining HIV-1 subtypes.

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BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates nucleotide sequences of the 5' adjacent region (C2 region) of the V3 region for the env gene in various subtypes of HIV-1. Capital letters indicate nucleotides that are entirely the same within a given subtype, and lower case letters indicate nucleotide variants within a given subtype. A question mark indicates that a consensus nucleotide was not determined because of too many variants. A dash indicates a nucleotide identical to that in subtype A. A period indicates the absence of a nucleotide in the corresponding site.

Figure 2 illustrates nucleotide sequences of the 3' adjacent region (C3 region) of the V3 region for the env gene in various subtypes of HIV-1. Capital letters indicate nucleotides that are entirely the same within a given subtype, and lower case letters indicate nucleotide variants within a given subtype. A question mark indicates that a consensus nucleotide was not determined because of too many variants. A dash indicates a nucleotide identical to that in subtype A. A period indicates the absence of a nucleotide in the corresponding site.

Figure 3 illustrates the locations, combinations, and base sequences of primers used in nested PCR

sequence GCAATAGAAAAATTCTCCTC (Sequence ID No. 5) and primer 12B containing nucleotide sequence ACAGTAGAAAAATTCCCCCTC (Sequence ID No. 6), and a mixture of primer 9AE containing nucleotide sequence

- 5 CACAGTACAATGCACACATG (Sequence ID No. 8) and primer 9B containing nucleotide sequence CACAGTACAATGTACACATG (Sequence ID No. 9), and using as the second primer pair primer 11VB containing nucleotide sequence CACAATTAAAACTGTGCATTAC (Sequence ID No. 28) and primer 10U
10 containing nucleotide sequence CTGTTAAATGGCAGTCTAGC (Sequence ID No. 20);

- (c) detecting subtype C using as the first primer pair a mixture of primer 12A containing nucleotide sequence GCAATAGAAAAATTCTCCTC (Sequence ID No. 5) and
15 primer 12B containing nucleotide sequence ACAGTAGAAAAATTCCCCCTC (Sequence ID No. 6), and a mixture of primer 9AE containing nucleotide sequence CACAGTACAATGCACACATG (Sequence ID No. 8) and primer 9B containing nucleotide sequence CACAGTACAATGTACACATG
20 (Sequence ID No. 9), and using as the second primer pair primer 11XC containing nucleotide sequence TTGTTTTATTAGGGAAGTGTTT (Sequence ID No. 29) and primer 10U containing nucleotide sequence CTGTTAAATGGTAGTCTAGC (Sequence ID No. 24); and

- 25 (d) detecting subtype C using as the first primer

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pair a mixture of primer 12A containing nucleotide
sequence GCAATAGAAAAATTCCTCCTC (Sequence ID No. 5) and
primer 12B containing nucleotide sequence
ACAGTAGAAAAATTCCTCCTC (Sequence ID No. 6), and a mixture of
5 primer 9AE containing nucleotide sequence
CACAGTACAATGCACACATG (Sequence ID No. 8) and primer 9B
containing nucleotide sequence CACAGTACAATGTACACATG
(Sequence ID No. 9), and using as the second primer pair
primer 11WE containing nucleotide sequence
10 CTCTACAATTAAATGATGCATTG (Sequence ID No. 30) and primer
10U containing nucleotide sequence CTGTTAAATGGCAGTCTAGC
(Sequence ID No. 20).

Alternatively, at least two subtypes can be
distinguished by repeating at least once the following
15 series of operations with different pairs of first and
second primers, where the operations comprise a first
amplification reaction that is carried out with a first
pair of primers using as a target sequence a portion of a
nucleotide sequence of the env gene of HIV-1, a second
20 amplification reaction that is then carried out with a
second pair of primers using as a target sequence a
nucleotide sequence within the target sequence in the
first reaction, and the detection of subtypes depending on
whether or not the nucleic acid has been amplified by the
25 second amplification reaction. For example, subtypes A, B,

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includes a sequence complementary to a portion of a nucleotide sequence in the C2 region of the env gene of HIV-1 and a primer (primer 3') which includes a sequence complementary to a portion of the nucleotide sequence of
5 the C3 region of the env gene for HIV-1 that differs depending on subtype (that is, subtype-specific nucleotide sequence). Since the C2 region of the env gene of HIV-1 has a nucleotide sequence that differs depending on the subtype, as shown in Figure 1, the nucleotide sequence may
10 be selected from this region to design primer 4'. Because the C3 region of the env gene of HIV-1 varies depending on the subtype, as shown in Figure 2, the nucleotide sequence may be selected from this region to design primer 3'. The primer length should generally be 18 to 30 base pairs, and
15 preferably 20 to 25 base pairs. The following primers can be specifically used.

Primer pairs and their nucleotide sequences for first PCR to detect subtype A

9AE/12A

- 20 primer 9AE: CACAGTACAATGCACACATG (Sequence ID No. 8)
primer 12A: GCAATAGAAAAATTCCTCCTC (Sequence ID No. 5)

Primer 9AE is a subtype A, E, F, and H-specific primer in which the sequence is complementary to the sequence from 6943 to 6962, counting from the 5' terminal
25 (left terminal), of the complete base sequence for HIV-1

primer 12E: GCAATAGAAAAATTCCCCTC (Sequence ID No. 7)

Primer 12E is a primer specific to subtype E only, in which the sequence is complementary to the sequence from 6943 to 6962, counting from the 5' terminal (left 5 terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer pairs and their nucleotide sequences for first PCR to detect subtype C

9B/12A

10 primer 9B: CACAGTACAATGTACACATG (Sequence ID No. 9)

primer 12A: GCAATAGAAAAATTCTCCTC (Sequence ID No. 5)

Primer pairs and their nucleotide sequences for first PCR to detect subtype D

9B/12B

15 primer 9B: CACAGTACAATGTACACATG (Sequence ID No. 9)

primer 12B: ACAGTAGAAAAATTCCCCTC (Sequence ID No. 6)

Primer pairs and their nucleotide sequences for first PCR to detect subtype F

9B/12A

20 primer 9B: CACAGTACAATGTACACATG (Sequence ID No. 9)

primer 12A: GCAATAGAAAAATTCTCCTC (Sequence ID No. 5)

Primer pairs and their nucleotide sequences for first PCR to detect subtype G

9B/12A

25 primer 9B: CACAGTACAATGTACACATG (Sequence ID No. 9)

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of the 3' adjacent region (C3 region) of the V3 region of the env gene for various subtypes of HIV-1. Since the nucleotide sequence varies depending on the subtype, a suitable sequence can be selected to design a primer. To
5 design a subtype-specific primer, phylogenetic analysis is employed to select nucleotide sequences of a given subtype which are as genetically remote as possible from the corresponding nucleotide sequence of other subtypes. An example can include a primer (primer 2) containing a
10 sequence complementary to a portion of the nucleotide sequence of the C3 region of the env gene for HIV-1. The primers pairs containing the following nucleotide sequences are specific examples.

Primer pairs and their nucleotide sequences for
15 second PCR to detect subtype A
10/11QA
primer 10: AAATGGCAGTCTAGCAGAAG (Sequence ID No. 4)
primer 11QA: CTCCTGAGGGGTTAGCAAAG (Sequence ID
No. 1)

20 Primer 10 is a subtype A, B, D, and E-specific primer in which the sequence is complementary to the sequence from 6997 to 7016, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

25 Primer 11QA is a primer specific to only subtype A,

in which the sequence is complementary to the sequence from 7313 to 7294, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

5 10U/11QA1

primer 10U: CTGTTAAATGGCAGTCTAGC (Sequence ID No. 20)

primer 11QA1: CTCCTGAGGAGTTAGCAAAG (Sequence ID No. 27)

Primer 10U is a subtype A, B, D, E, and J-specific primer in which the sequence is complementary to the
10 sequence from 6992 to 7011, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11QA1 is a primer specific to only subtype A, in which the sequence is complementary to the sequence
15 from 7313 to 7294, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer pairs and their nucleotide sequences for second PCR to detect subtype B

20 10/11BB

primer 10: AAATGGCAGTCTAGCAGAAG (Sequence ID No. 4)

primer 11BB: CTGTGCATTACAATTTCTGG (Sequence ID No. 2)

Primer 11BB is a primer specific to only subtype B,
25 in which the sequence is complementary to the sequence

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Primer 11WE is a primer specific to only subtype E, in which the sequence is complementary to the sequence from 7352 to 73339, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer pairs and their nucleotide sequences for second PCR to detect subtype C

10C/11RC

primer 10C: AAATGGTAGCCTAGCAGAAG (Sequence ID No. 10)
10 primer 11RC: CTCCTGAGGATGGTGCAAATTT (Sequence ID No. 13)

Primer 10C is a subtype C and F-specific primer in which the sequence is complementary to the sequence from 6997 to 7016, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11RC is a primer specific to only subtype C, in which the sequence is complementary to the sequence from 7313 to 7292, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

10U/11XC

primer 10U: AAATGGTAGCCTAGCAGAAG (Sequence ID No. 10)
primer 11XC: TTGTTTATTAGGGAAGTGTTTC (Sequence ID No. 29)

Primer 11XC is a primer specific to only subtype C, in which the sequence is complementary to the sequence

from 7289 to 7268, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

- Primer pairs and their nucleotide sequences for
5 second PCR to detect subtype D

10/11RD

primer 10: AAATGGCAGTCTAGCAGAAG (Sequence ID No. 4)

primer 11RD: CTCCTGAGGATGGTTAAAAAT (Sequence ID No. 14)

- Primer 11RD is a primer specific to only subtype D,
10 in which the sequence is complementary to the sequence
from 7313 to 7292, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

- Primer pairs and their nucleotide sequences for
15 second PCR to detect subtype F

10C/11RF

primer 10C: AAATGGTAGCCTAGCAGAAG (Sequence ID No. 10)

primer 11RF: CTCCTGAGGATGAGTTAAATTT (Sequence ID No. 15)

- Primer 11RF is a primer specific to only subtype F,
20 in which the sequence is complementary to the sequence
from 7313 to 7292, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

- Primer pairs and their nucleotide sequences for
25 second PCR to detect subtype G

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10G/11SG

primer 10G: GAATGGCAGTTTAGCAGAAG (Sequence ID No. 11)

primer 11SG: TCCTGCAGATGAGTTAAAGG (Sequence ID No. 16)

Primer 10G is a primer specific to only subtype G,
5 in which the sequence is complementary to the sequence
from 6997 to 7016, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

Primer 11SG is a primer specific to only subtype G,
10 in which the sequence is complementary to the sequence
from 7312 to 7293, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

Primer pairs and their nucleotide sequences for
15 second PCR to detect subtype H

10H/11SH

primer 10H: GTCAAATGGCAGTTTAGCAG (Sequence ID No. 12)

primer 11SH: TCCTGAGGATGGTTTAAAGG (Sequence ID No. 17)

Primer 10H is a primer specific to only subtype H,
20 in which the sequence is complementary to the sequence
from 6994 to 7013, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

Primer 11SH is a primer specific to only subtype H,
25 in which the sequence is complementary to the sequence

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from 7312 to 7293, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Second PCR may alternatively be carried out using a mixture of primers capable of giving amplified products for several subtypes in order to permit the amplification of any subtype. Examples of primers for that purpose include the following primer mixtures.

- primer 10U: AATGGCAGTCTAGCAGAAG (Sequence ID No. 4)
10 primer 11LB: AATTTCTGGGTCCCCTCCTG (Sequence ID No. 18)
primer 11LAE: AATTTCTAGATCCCCTCCTG (Sequence ID No. 25)
primer 11LC: AATTTCTAGGTCCCCTCCTG (Sequence ID No. 26)

Primer 11LB is a subtype B, D, F, G, and I-specific primer in which the sequence is complementary to the sequence from 7327 to 7308, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11LAE is a subtype A, E, F, G, I, and J-specific primer in which the sequence is complementary to the sequence from 7327 to 7308, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11LC is a subtype C, F, G, H, I, and J-specific primer in which the sequence is complementary to

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6992 to 7011, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11LE: AATTCTCTAGATCTCCTCCTG)Sequence ID No. 19)

- 5 Primer 11LE is a subtype E, F, G, H, and J-specific primer in which the sequence is complementary to the sequence from 7327 to 7308, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

- 10 Primer 11LC: AATTCTCTAGGTCCTCCTG (Sequence ID No. 26)

Primer 11LC is a subtype C, F, G, H, I, and J-specific primer in which the sequence is complementary to the sequence from 7327 to 7308, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11TC: TTCTCCTCTACAATTAAAGC (Sequence ID No. 31)

- 20 Primer 11TC is a primer specific to only subtype C, in which the sequence is complementary to the sequence from 7357 to 7338, counting from the 5' terminal (left terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11RC1: TTATTGTTTTATTAGGGAAGTG (Sequence ID No. 32)

- 25 Primer 11RC1 is a primer specific to only subtype C, in which the sequence is complementary to the sequence from 7292 to 7271, counting from the 5' terminal (left

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terminal), of the complete base sequence for HIV-1 (NL4-3 strain).

Primer 11SE: TGCATTGTAATTTCTAGATCTC (Sequence ID No. 33)

Primer 11SE is a primer specific to only subtype E,
5 in which the sequence is complementary to the sequence
from 7333 to 7314, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

Primer 11BE: TGATGCATTGTAATTTCTAG (Sequence ID No. 34)

10 Primer 11BE is a primer specific to only subtype E,
in which the sequence is complementary to the sequence
from 7338 to 7319, counting from the 5' terminal (left
terminal), of the complete base sequence for HIV-1 (NL4-3
strain).

15 The PCR procedures and reaction conditions may be in
accordance with those in Bruisten S. et al., *AIDS Res Hum
Retroviruses* 1993, 9:259-265, but the hot start method is
preferred. In hot start PCR, the PCR reaction solution is
kept on a hot plate for start up at an elevated
20 temperature (usually 90°C or more).

However, it sometimes happens that no subtype is
detected in attempts to determine the HIV-1 subtype in
such a method. Possible causes may be that the HIV-1 DNA
concentration is below the detection threshold, or the
25 presence of numerous variants at the primer binding site.

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To deal with the former possibility, the above method can be implemented after the extraction of the RNA from plasma, since the concentration of HIV-1 is generally higher in plasma than in cells, and its subsequent
5 conversion to DNA using reverse transcriptase.

To deal with the latter possibility, the determination of the subtype by this method is held off, another genetic region of HIV-1 is amplified by PCR to determine the nucleotide sequence, and the subtype is
10 determined by a conventional method (Note: HIV-1 infection is generally diagnosed by detecting antibodies. This invention is not a method for diagnosing HIV-1 infection.).

The PCR reaction products are separated by agarose gel electrophoresis and detected by ethidium bromide
15 staining. Although distinct bands can be observed with the use of primers consistent with the subtype of the HIV-1 in sample DNA, the bands are indistinct or not observed at all when the primers are not. The HIV-1 subtype is determined in this way.

Determining the HIV-1 subtype in the present invention may include the steps of amplifying nucleic acid using as a target sequence a portion of a nucleotide
sequence of the HIV-1 genome, where the nucleotide sequence is highly conserved among all subtypes, and
25 determining the presence of absence of HIV-1 depending on

caused by inconsistencies between the primer and analyte HIV-1 DNA base pairs, the magnesium ion concentration of the reaction buffer should be increased from the usual concentration of 1.5 mM to 4 mM.

- 5 The components constituting the diagnostic kit may be packaged individually, assembled, or bundled in containers such as vials and tubes, and the containers may be bundled and housed in support means divided for housing such components.

10 BEST MODE FOR CARRYING OUT THE INVENTION

The present invention is illustrated in detail in the following examples, but the scope of the present invention is not in any way limited by these examples.

Example 1

15 Subjects and Methodology

1) Subtype-specific reference specimens to study method for determining subtype

Reference specimens were prepared after the extraction of DNA from the blood of 3, 8, and 3 HIV-1-
20 infected patients; the subtypes of HIV-1 were determined to be A, B, and E, respectively, by env gene sequencing and phylogenetic analysis.

2) Subjects for determining subtype

The HIV-1 subtype was determined in 8 subjects with
25 HIV who either visited or were hospitalized in Tokyo

hospitals.

3) Preparation of DNA from blood of HIV patients

10 mL peripheral blood was drawn from the above HIV patients. Sodium citrate was used as an anticoagulant.

- 5 Monocytes were separated from the peripheral blood by Ficoll-Paque (Pharmacia) gradient centrifugation, and DNA was then prepared using a QIAamp Blood Kit (QIAGEN). The DNA was dissolved in pure water or buffer containing 1 mM EDTA, and was stored at -20°C until use. 0.5 µg DNA was
10 used in PCR.

4) Detection of subtypes A, B, and E by PCR

Figure 3 gives the nucleotide sequences of the primers used in PCR.

- For subtype A-specific detection, nested PCR was
15 carried out using 9AE and 12A as the primers for first PCR, and 10 and 11QA as the primers for second PCR. For subtype B-specific detection, nested PCR was carried out using 9B and 12B as the primers for first PCR, and 10 and 11BB as the primers for second PCR. For subtype E-
20 specific detection, nested PCR was carried out using 9AE and 12E as the primers for first PCR, and 10 and 11QE as the primers for second PCR (Figure 3).

- PCR was carried out for 30 cycles, where one cycle was 15 seconds at 94°C, 30 seconds at 56°C, and 1 minute
25 at 72°C, with 100 µL reaction solution (10 mM Tris-HCl pH

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8.3, 50 mM KCl, 4 mM MgCl₂, 0.2 mM dNTP, 1.0 μM primer, 2.5 units Taq polymerase) using 0.5 μg sample DNA prepared from HIV patients. Second PCR was carried out for 25 cycles under the same conditions using 2 μL reaction solution from the first PCR. 30 seconds at 60°C was used instead of 30 seconds at 56°C, however.

PCR products (subtypes A and E: 317 bp; subtype B: 342 bp) were detected by isolation through electrophoresis with 2% agarose gel, and by subsequent ethidium bromide staining.

Results

1) Study of subtype determination by PCR of subtype-specific reference samples

The following results were obtained for specimens whose subtype had been determined by virus genome sequencing. Only subtype A specimens were positive, while subtype B and E specimens were all negative, in PCR using primers for the detection of subtype A. Only subtype B specimens were positive, while subtype A and E specimens were all negative, in PCR using primers for the detection of subtype B. Only subtype E specimens were positive, while subtype A and B specimens were all negative, in PCR using primers for the detection of subtype E (Figure 4).

2) Determination of HIV patient subtype by PCR

Table 1 gives the results obtained in the

method was able to correctly diagnose the subtype in 21
out of 22 cases. The determination was postponed in the
remaining one case. That is, the present method has been
shown to be a simple and reliable method for determining
5 subtypes.

The method of the present invention allows HIV-1
subtypes to be determined at a cost of about ¥2,000 per
specimen. The time needed to determine the subtype in
treating all 8 specimens at once was 2 hour for the
10 isolation of the DNA, 6 hours for PCR, and about 1 hour
for electrophoresis.

Example 2

Subjects and Methodology

- 1) Subtype-specific reference specimens for
15 detection in subtype determination method

Reference specimens were prepared from DNA extracted
from the blood of 11 subjects, which included 2 patients
with HIV-1 determined to be subtype C by env gene
sequencing and phylogenetic analysis, in addition to the 3
20 subtype A HIV-1 subjects, 3 subtype B HIV-1 subjects, and
3 subtype E HIV-1 subjects used in Example 2.

- 2) Subjects for determining subtype

The HIV-1 subtype was determined in 32 subjects with
HIV who either visited or were hospitalized in Tokyo
25 hospitals.

3) Preparation of DNA from blood of HIV patients

10 mL peripheral blood was drawn from the above HIV patients. Sodium citrate was used as an anticoagulant. Monocytes were separated from the peripheral blood by Ficoll-Paque (Pharmacia) gradient centrifugation, and DNA was then prepared using a QIAamp Blood Kit (QIAGEN). The DNA was dissolved in pure water or buffer containing 1 mM EDTA, and was stored at -20°C until immediately before use. 0.5 µg DNA was used in PCR.

4) Detection of subtypes A, B, C, and E by PCR

Figure 5 gives the nucleotide sequences of the primers used in PCR.

A mixture of 9AE, 9B, 12A, and 12B was used for the primers in first PCR. Nested PCR was carried out using the following primers for second PCR: 10U and 11QA1 for subtype A-specific detection, 10U and 11VB for subtype B-specific detection, 10U and 11XC for subtype C-specific detection, and 10U and 11WE for subtype E-specific detection. Nested PCR was carried out using a mixture of 10U, 11LB, 11LAE, and 11LC for amplification of HIV-1 DNA irrespective of subtype (Figure 5).

PCR was carried out for 30 cycles, where one cycle was 15 seconds at 94°C, 30 seconds at 56°C, and 1 minute at 72°C, with 100 µL reaction solution (10 mM Tris-HCl pH 8.3, 50 mM KCl, 4 mM MgCl₂, 0.2 mM dNTP, 1.0 µM primer,

2.5 units Taq polymerase) using 0.5 µg sample DNA prepared from HIV patients. Second PCR was carried out for 25 cycles under the same conditions using 2 µL reaction solution from the first PCR. 30 seconds at 60°C was used instead of 30 seconds at 56°C, however.

PCR products (subtype A: 322 bp; subtype B: 358 bp; subtype C: 298 bp; subtype E: 361)) were detected by isolation through electrophoresis with 2% agarose gel, and by subsequent ethidium bromide staining.

10 Results

1) Study of subtype by PCR of subtype-specific reference samples

The following results were obtained for specimens whose subtype had been determined by virus genome sequencing. Only subtype A specimens were positive, while subtype B, C, and E specimens were all negative, in PCR using primers for the detection of subtype A. Only subtype B specimens were positive, while subtype A, C, and E specimens were all negative, in PCR using primers for the detection of subtype B. Only subtype C specimens were positive, while subtype A, B, and E specimens were all negative, in PCR using primers for the detection of subtype C. Only subtype E specimens were positive, while subtype A, B, and C specimens were all negative, in PCR using primers for the detection of subtype E (Figure 6).

All specimens were positive in PCR using primers for amplifying HIV-1 DNA irrespective of subtype (Figure 6).

2) Determination of subtype of HIV patients by PCR

The HIV-1 subtype was determined in 32 HIV patients who either visited or were hospitalized in Tokyo hospitals. Together with the 11 subtype-specific reference samples, there were 3 subtype A cases, 30 subtype B cases, 2 subtype C cases, and 8 subtype E cases.

To verify that the above results were correct, the amplified DNA of 21 out of the 43 HIV patients whose subtype was determined by PCR was sequenced and phylogenetically analyzed, giving the results shown in Figure 7. There was complete agreement between the subtypes determined by phylogenetic analysis and the subtypes determined by PCR for these cases of HIV-1.

The method employed in Example 2 differs significantly from that of Example 1 in that universal primers were used in first PCR, and subtype-specific primers were used in second PCR. As a result, the number of PCR reactions could be reduced to 5/8. In view of the fact that the determination of the subtype had to be postponed in 1 case in Example 1, the method of Example 2 was able to provide more accurate diagnosis and was also simpler.

3) Effect of subtype on testing for drug resistance

by genotype

Drug resistance by genotype was analyzed based on data for subtype B. To investigate whether or not the drug resistance of HIV-1 subtypes other than subtype B could be determined using data for subtype B, the amino acid sequences for HIV-1 protease before and after HAART treatment were determined in 4 patients infected with HIV-1 other than subtype B who were receiving HAART treatment. After HAART treatment in case C3, which was subtype E, amino acid No. 10 had mutated from L (leucine) to F (phenylalanine), and amino acid No. 20 had mutated from K (lysine) to T (threonine). This was recognized as an amino acid mutation indicative of drug resistance in subtype B. However, in all four subjects, amino acid No. 36 was I (isoleucine) from before HAART treatment. In the data for subtype B, HIV-1 with I as the No. 36 amino acid was interpreted as indicative of drug resistance. However, it is difficult to conclude that HIV-1 would have acquired drug resistance before administration of the drug. It would be more logical to view this mutation as irrelevant to drug resistance in HIV-1 subtypes other than subtype B. It may thus be concluded that it is important to diagnose the subtype in advance in order to properly assess drug resistance by genotype.

4) Relationship between subtype and sexual behavior

Figure 9 summarizes the relation between subtype and sexual behavior in 22 patients with HIV who had been interviewed about their sexual preferences. Heterosexuals are those attracted to the opposite sex, while MSM are male homosexuals. About the same number of heterosexuals were subtype B and E, whereas male homosexuals were far more likely to be subtype B. This would seem to indicate that Southeast Asian HIV has spread among heterosexuals but has not spread very much among male homosexuals.

10 Example 3

Subjects and Methodology

1) Western blot-negative and PA-positive serum specimens

The specimens were 15 serum samples whose blood tests at Tokyo hospitals showed to be HIV-1 negative by Western blotting and HIV-1 positive by PA.

2) Preparation of DNA from plasma RNA

RNA was prepared using an RNAeasy Kit (QIAGEN) from 200 μ L of the aforementioned serum specimen. The RNA was dissolved in pure water and stored at -20°C until immediately before use.

3) Detection of HIV-1 by PCR

RNA corresponding to 20 μ L portions of serum was used as the material, and cDNA was synthesized by 30 minutes of reaction at 42°C using a mixture of primers 12A

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and 12B with 20 μ L reaction solution (10 mM Tris-HCl pH 8.3, 50 mM KCl, 2.5 mM $MgCl_2$, 1 mM dNTP, 5 μ M primer, and 100 unites reverse transcriptase). The cDNA was used as material in nested PCR capable of amplifying the DNA of HIV-1 irrespective of subtype. A mixture of 9AE, 9B, 12A, and 12B was used for the primers in first PCR. A mixture of 10U, 11LB, 11LAE, and 11LC was used for the primers in second PCR.

First PCR was carried out for 30 cycles, where one cycle was 15 seconds at 94°C, 30 seconds at 56°C, and 1 minute at 72°C, with 100 μ L reaction solution (10 mM Tris-HCl pH 8.3, 50 mM KCl, 4 mM $MgCl_2$, 0.2 mM dNTP, 1.0 μ M primer, 2.5 units Taq polymerase). Second PCR was carried out for 25 cycles under the same conditions using 2 μ L reaction solution from the first PCR. 30 seconds at 60°C was used instead of 30 seconds at 56°C, however.

PCR products (subtype A: 322 bp; subtype B: 358 bp; subtype C: 298 bp; subtype E: 361) were detected by isolation through electrophoresis with 2% agarose gel, and by subsequent ethidium bromide staining.

Results

No HIV-1 was detected by PCR in any of the 15 serum specimens which were HIV-1 negative by Western blotting and HIV-1 positive by PA (Figure 10). It was proven (Figure 6) that the PCR run here was designed to enable

primer 12E.

Sequence ID No. 8 gives the nucleotide sequence for
primer 9AE.

Sequence ID No. 9 gives the nucleotide sequence for
5 primer 9B.

Sequence ID No. 10 gives the nucleotide sequence for
primer 10C.

Sequence ID No. 11 gives the nucleotide sequence for
primer 10G.

10 Sequence ID No. 12 gives the nucleotide sequence for
primer 10H.

Sequence ID No. 13 gives the nucleotide sequence for
primer 11RC.

Sequence ID No. 14 gives the nucleotide sequence for
15 primer 11RD.

Sequence ID No. 15 gives the nucleotide sequence for
primer 11RF.

Sequence ID No. 16 gives the nucleotide sequence for
primer 11SG.

20 Sequence ID No. 17 gives the nucleotide sequence for
primer 11SH.

Sequence ID No. 18 gives the nucleotide sequence for
primer 11LB.

Sequence ID No. 19 gives the nucleotide sequence for
25 primer 11LE.

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CLAIMS

1. A method for determining HIV-1 subtypes, characterized by comprising the steps of amplifying nucleic acid using as a target sequence a portion of a
5 nucleotide sequence of the env gene of HIV-1, where at least one of the 5' terminal and 3' terminal nucleotide sequences is different depending on the HIV-1 subtype, and detecting the subtype depending on whether or not the nucleic acid has been amplified.

10 2. The method according to Claim 1, wherein the target sequence is 100 to 2500 nucleotides long.

3. The method according to Claim 1, wherein the sequence from the 1st through 30th bases from the 3' terminal and/or 5' terminal of the target sequence is
15 different depending on the subtype.

4. The method according to Claim 3, wherein the 3' terminal of the target sequence is in the C3 region of the env gene of HIV-1.

5. The method according to Claim 4, wherein the
20 5' terminal of the target sequence is in the C2 region of the env gene of HIV-1.

6. The method according to Claim 1, wherein different amplification reactions are carried out using different pairs of primers, and different subtypes are
25 detected.

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nucleotide sequence (nucleotide sequence 1) that differs depending on subtype in the C3 region of the env gene of HIV-1, and a primer (primer 2) that includes a sequence complementary to a portion of the nucleotide sequence (nucleotide sequence 2) of the C2 region of the env gene of HIV-1; and the first pair of primers consists of a primer (primer 3) that includes a sequence complementary to a portion of a nucleotide sequence (nucleotide sequence 3) of a region downstream of the 3' terminal of nucleotide sequence 1 of the env gene of HIV-1, and a primer (primer 4) that includes a sequence complementary to a portion of a nucleotide sequence (nucleotide sequence 4) of a region upstream of the 5' terminal of nucleotide sequence 2 of the env gene of HIV-1.

10. The method according to Claim 8, wherein at least two subtypes are distinguished by repeating at least once, with different pairs of second primers, a series of operations comprising: a first amplification reaction that is carried out with the first pair of primers using as a target sequence a portion of a nucleotide sequence of the env gene of HIV-1; a second amplification reaction that is then carried out with the second pair of primers using as a target sequence a nucleotide sequence within said target sequence; and the detection of subtypes depending on whether or not the nucleic acid has been amplified by the

second amplification reaction.

11. The method according to Claim 10, wherein subtypes A, B, C, and E are distinguished by:

- (a) detecting subtype A using as the first primer
- 5 pair a mixture of primer 12A containing nucleotide sequence GCAATAGAAAAATTCTCCTC (Sequence ID No. 5) and primer 12B containing nucleotide sequence ACAGTAGAAAAATCCCCCTC (Sequence ID No. 6), and a mixture of primer 9AE containing nucleotide sequence
- 10 CACAGTACAATGCACACATG (Sequence ID No. 8) and primer 9B containing nucleotide sequence CACAGTACAATGTACACATG (Sequence ID No. 9), and using as the second primer pair primer 11QA1 containing nucleotide sequence CTCCTGAGGAGTTAGCAAAG (Sequence ID No. 27) and primer 10U
- 15 containing nucleotide sequence CTGTTAAATGGCAGTCTAGC (Sequence ID No. 20);

- (b) detecting subtype B using as the first primer pair a mixture of primer 12A containing nucleotide sequence GCAATAGAAAAATTCTCCTC (Sequence ID No. 5) and
- 20 primer 12B containing nucleotide sequence ACAGTAGAAAAATCCCCCTC (Sequence ID No. 6), and a mixture of primer 9AE containing nucleotide sequence CACAGTACAATGCACACATG (Sequence ID No. 8) and primer 9B containing nucleotide sequence CACAGTACAATGTACACATG
- 25 (Sequence ID No. 9), and using as the second primer pair

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(Sequence ID No. 9), and using as the second primer pair primer 11WE containing nucleotide sequence CTCTACAATTAAATGATGCATTG (Sequence ID No. 30) and primer 10U containing nucleotide sequence CTGTTAAATGGCAGTCTAGC (Sequence ID No. 20).

12. The method according to Claim 8, wherein at least two subtypes are distinguished by repeating at least once, with different pairs of first and second primers, a series of operations comprising: a first amplification reaction that is carried out with a first pair of primers using as a target sequence a portion of a nucleotide sequence of the env gene of HIV-1; a second amplification reaction that is then carried out with a second pair of primers using as a target sequence a nucleotide sequence within said target sequence; and the detection of subtypes depending on whether or not the nucleic acid has been amplified by the second amplification reaction.

13. The method according to Claim 12, wherein subtypes A, B, and E are distinguished by:

(a) detecting subtype A using as the first primer pair primer 12A containing nucleotide sequence GCAATAGAAAAATTCTCTCTC (Sequence ID No. 5) and primer 9AE containing nucleotide sequence CACAGTACAATGCACACATG (Sequence ID No. 8), and using as the second primer pair primer 11QA containing nucleotide sequence

CTCCTGAGGGGTTAGCAAAG (Sequence ID No. 1) and primer 10 containing nucleotide sequence AAATGGCAGTCTAGCAGAAG (Sequence ID No. 4);

- (b) detecting subtype B using as the first primer pair primer 12B containing nucleotide sequence ACAGTAGAAAAATTCCTC (Sequence ID No. 6) and primer 9B containing nucleotide sequence CACGTACAATGTACACATG (Sequence ID No. 9), and using as the second primer pair primer 11BB containing nucleotide sequence CTGTGCATTACAATTTCTGG (Sequence ID No. 2) and primer 10 containing nucleotide sequence AAATGGCAGTCTAGCAGAAG (Sequence ID No. 4); and

- (c) detecting subtype E using as the first primer pair primer 12E containing nucleotide sequence GCAATAGAAAAATTCCTC (Sequence ID No. 7) and primer 9AE containing nucleotide sequence CACGTACAATGCACACATG (Sequence ID No. 8), and using as the second primer pair primer 11QE containing nucleotide sequence CTCCTGAGGGTGGTTGAAAG (Sequence ID No. 3) and primer 10 containing nucleotide sequence AAATGGCAGTCTAGCAGAAG (Sequence ID No. 4).

14. The method according to Claim 1, further comprising the steps of amplifying nucleic acid using as a target sequence a portion of a nucleotide sequence of the HIV-1 genome, the nucleotide sequence being highly

conserved among all subtypes, and ascertaining the presence or absence of HIV-1 depending on whether or not the nucleic acid has been amplified.

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15. The method according to Claim 14, wherein the
5 step for ascertaining the presence or absence of HIV-1
comprises amplifying the nucleic acid with a first primer
pair using as a target sequence a portion of a nucleotide
sequence of the HIV-1 genome, the nucleotide sequence
being highly conserved among all subtypes, then carrying
10 out a second amplifying reaction with a second primer pair
using as a target sequence a nucleotide sequence in said
target sequence, and ascertaining the presence or absence
of HIV-1 depending on whether or not the nucleic acid has
been amplified.

15 16. The method according to Claim 15, wherein the
primers that are used comprise a mixture of a plurality of
upstream primers with differing nucleotide sequences and a
plurality of downstream primers with differing nucleotide
sequences.

20 17. The method according to Claim 16, wherein the
first primers comprise a mixture of primer 12A containing
nucleotide sequence GCAATAGAAAAATTCTCCTC (Sequence ID
No. 5), primer 12B containing nucleotide sequence
ACAGTAGAAAAATTCCTCCTC (Sequence ID No. 6), primer 9AE
25 containing nucleotide sequence CACAGTACAATGCACACATG

(Sequence ID No. 8), and primer 9B nucleotide sequence CACAGTACAATGTACACATG (Sequence ID No. 9), and the second primer pair comprises primer 11LB containing nucleotide sequence AATTTCTGGGTCCCTCCTG (Sequence ID No. 18), primer
5 11LAE containing nucleotide sequence AATTTCTAGATCCCTCCTG (Sequence ID No. 25), primer 11LC containing nucleotide sequence AATTTCTAGGTCCCTCCTG (Sequence ID No. 26), and primer 10U containing nucleotide sequence CTGTTAAATGGCAGTCTAGC (Sequence ID No. 20).

- 10 18. A kit for determining HIV-1 subtypes, comprising primer pairs in which a target sequence is a portion of a nucleotide sequence of the env gene of HIV-1, where at least one of the 5' terminal and 3' terminal nucleotide sequences is different depending on the subtype.

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Fig. 1

SUBTYPE A TGTaataccTCagccattACacAgGCTGTGCCaAAggTatCCTTTGAgCCaATTCCCATA
 SUBTYPE B -----c-----t-----c-----
 SUBTYPE C -----c-----t-----A-----a-----C-----c-----t-----c-----T-----
 SUBTYPE D -----g-----a-----a-----
 SUBTYPE E -----T-----T-----aG-----a-----T-----t-----
 SUBTYPE F -----A-----T-----GG-----T-----
 SUBTYPE G -----gt-----A-----A-----ga?T-----c-----
 SUBTYPE H -----GT-----A-----GAGT-----T-----

SUBTYPE A caTTATGTGtGcCCagGCTGgtTttGCgATtCTAAagTGtAa?gataaggagTTcaatGGA
 SUBTYPE B -----g-----t-----a-----
 SUBTYPE C -----t-----a-----ta-----aca-----g
 SUBTYPE D -----a-----a-----a-----A-----g
 SUBTYPE E -----a-----t-----a-----t-----T-----a-----t-----g
 SUBTYPE F -----T-----A-----T-----aA-----G
 SUBTYPE G -----T-----t-----gg-----a-----?
 SUBTYPE H -----T-----G-----A-----GG-----A-----G

SUBTYPE A acAGGgccatGcaagAATGTcAGcaCaGTaCAATGcACacATGGaATcAagCCAGtagTa
 SUBTYPE B -----a-----t-----ca-----t-----t-----g-----
 SUBTYPE C -----a-----c-----t-----t-----t-----g-----
 SUBTYPE D -----?-----a-----t-----g-----t-----g-----
 SUBTYPE E -----t-----A-----T-----T-----T-----G-----
 SUBTYPE F -----g-----T-----T-----T-----A-----g-----
 SUBTYPE G -----A-----T-----a-----T-----T-----T-----g-----
 SUBTYPE H -----G-----AA-----T-----A-----T-----T-----T-----G-----

SUBTYPE A tCAACTCAaCTGcTGtTaAATGGcAGtcTAGCAGAgaAa???gaggTAatgaTtagaTCT
 SUBTYPE B -----a-----t-----c-----g-----a-----
 SUBTYPE C -----?-----g-----a-----A-----
 SUBTYPE D -----t-----t-----C-----ta-----A-----C-----
 SUBTYPE E -----T-----T-----C-----ta-----A-----C-----
 SUBTYPE F -----t-----a-----c-----g-----t-----aA-----a-----c-----
 SUBTYPE G -----T-----A-----C-----GTCAAATG-----CAGTTT-----C-----?-----a-----a-----

SUBTYPE A gAaaataTcacAaAaATgccaaaAccaTAaTaGTacAgcTtg???aagcctGTAA?aAtt
 SUBTYPE B -----t-----gg-----t-----gaa-----g-----at-----ga-----
 SUBTYPE C -----c-----g-----t-----a-----t-----aAtg-----at-----ga-----
 SUBTYPE D -----c-----c-----t-----?-----AAtG-----t-----?-----c-----
 SUBTYPE E -----C-----g-----t-----A-----G-----C-----AAT-----At-----Ga-----C-----
 SUBTYPE F -----c-----t-----g-----t-----A-----?-----AATg-----At-----ca-----
 SUBTYPE G -----c-----g-----g-----gt-----g-----AAt-----a-----a-----ga-----
 SUBTYPE H -----c-----g-----a-----gt-----AAt-----a-----?-----g-----

SUBTYPE A aatTGT
 SUBTYPE B -----
 SUBTYPE C gtg-----
 SUBTYPE D -----
 SUBTYPE E -----
 SUBTYPE F -----
 SUBTYPE G --?--
 SUBTYPE H --?--

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Fig. 2

SUBTYPE A TGTAatgTcAgtaga?cagaaTGGaAtaaaacttTacaa?aggtagcta?acAatTAaga
 SUBTYPE B -----ca-t-----g-a-----c-----a-c-a-t-----A-----
 SUBTYPE C -----CA-T-----a-ga?a-----?------a-----ag-a-a-----gc-----
 SUBTYPE D -----a-T-----a-ga?a-----c-----a-a-----g-----
 SUBTYPE E -----G-gA-T-A-g-A-a-----g-g-----a-c-----a-ga-a-----a-----
 SUBTYPE F -----c-t-----g-a-C-----?-?-g-a-----a?ggc-a-g-----ag-----
 SUBTYPE G -----t-----a?-a-t-----?g-g-tG-----ga-t-----a??gc-?-C-----a-----
 SUBTYPE H -----T-----g?-a-?------g-g-tg-----?-a-----?-?-c-----a-----

SUBTYPE A aaa.....tacTtt?????????aacaaaaa...?????ataatcTTtgctaac...
 SUBTYPE B g-??????c-a...g-g-t-----g-----aa-C-a???
 SUBTYPE C g-.....c-----ccct-----T-----aa-----acca...
 SUBTYPE D g-c?.....cTto-----aca-----t-----aaacCa...
 SUBTYPE E g-g.....C-----a-t-T-G-----caaCCA???
 SUBTYPE F tct.....c-t-c-----tg-----aa-----aactcA...
 SUBTYPE G g-.....at-----?c???-----c-----aaCtCA...
 SUBTYPE H ---.....?-----a-----t-----??-----c-----aaacca???

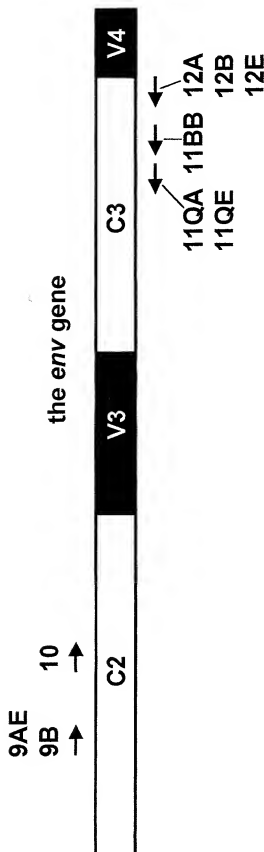
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 SUBTYPE B t-----cCc-----gt-tg-c-----g-----
 SUBTYPE C t-----Cc-----c-----a-----g-----
 SUBTYPE D t-----ccc-----c-----g-----
 SUBTYPE E c-----a-C-----tg-ca-----A-----g-----
 SUBTYPE F t-----CC-----tg-----a-----g-----
 SUBTYPE G t-tg-----cC-----a-----
 SUBTYPE H t-----Cc-----?-a-----

SUBTYPE A TAtTgc
 SUBTYPE B --c-t
 SUBTYPE C -----
 SUBTYPE D --C-----
 SUBTYPE E -----
 SUBTYPE F --C-----
 SUBTYPE G -----t
 SUBTYPE H -----t

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Fig. 3



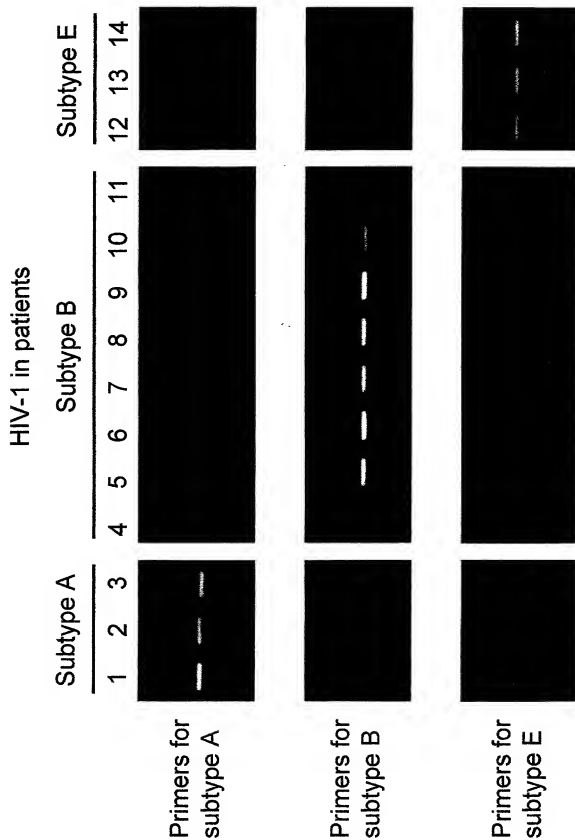
Primers

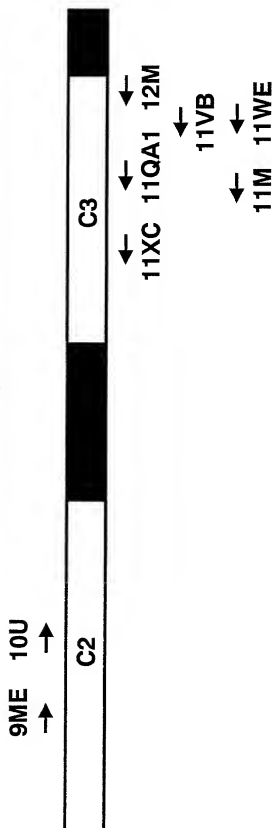
9AE: CACAGTACAATGCACACATG
 9B: CACAGTACAATGTACACATG
 10: AAATGGCAGTCTAGCAGAAG
 11QA: CTCCTGAGGGGTAGCAAAG
 11QE: CTCCTGAGGGGTGTTGAAAG
 11BB: CTGTGCATTACAATTTCTGG
 12A: GCAATAGAAAAAATTCCTCTC
 12B: ACAGTAGAAAAAATTCCTCTC
 12E: GCAATAGAAAAAATTCCTCTC

Primers used for		
Subtype	1st PCR	2nd PCR
A	9AE/12A	10/11QA
B	9B/12B	10/11BB
E	9AE/12E	10/11QE

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Fig. 4



the *env* gene

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Fig. 5

Primers used for		
Subtype	1st PCR	2nd PCR
A	9M/12M	10U/11QA1
B	9M/12M	10U/11VB
C	9M/12M	10U/11XC
E	9M/12M	10U/11WE
All	9M/12M	10U/11M

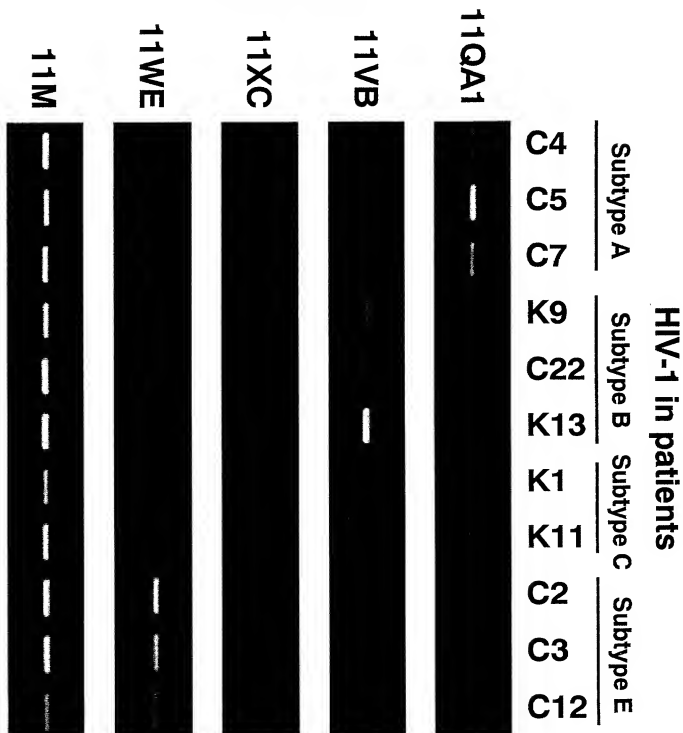
Primers

9AE: CACAGTACAATGCACACATG
 9B: CACAGTACAATGTACACATG
 10U: CTGTAAATGGCAGTCTAGC
 11QA1: CTCCTGAGGAGTTAGCAAAG
 11VB: CACAATTAAAACTGTGCATTAC
 11XC: TTGTTTTATTAGGAAAGTGTTT
 11WE: CTCACAAATTAATGATGCATTG
 11LAE: AATTCTAGATCCCTCCTG
 11LB: AATTCTGGTCCCTCCTG
 11LC: AATTCTAGTCCCTCCTG
 12A: GCAATAGAAAAATTCCTC
 12B: ACAGTAGAAAAATTCCTC

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Fig. 6

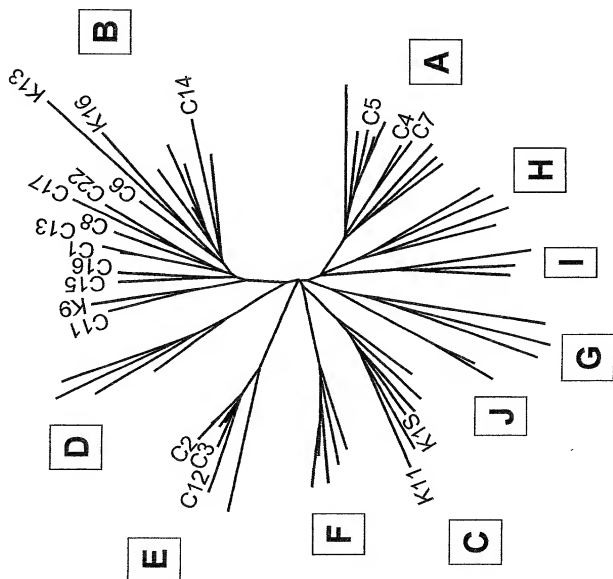
Primers used



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Fig. 7



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Fig. 8

Patient	Subtype	Date	Amino acid at positions relevant to PRI resistance															
			10	20	30	36	46	48	50	63	82	84	90					
C3	E	8/11/97	L	K	D	I	M	G	I	L	V	I	L					
		6/17/99	F	T	D	I	M	G	I	L	V	I	L					
C4	A	6/9/97	L	I	D	I	M	G	I	N	V	I	L					
		3/11/98	L	I	D	I	M	G	I	N	V	I	L					
C5	A	6/23/97	L	I	D	I	M	G	I	P	V	I	L					
		1/11/99	L	I	D	I	M	G	I	P	V	I	L					
C7	A	7/29/97	L	I	D	I	M	G	I	N	V	I	L					
		9/16/99	L	I	D	I	M	G	I	N	V	I	L					

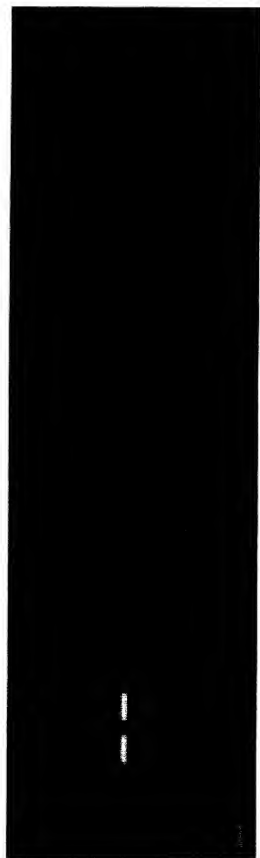
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Fig. 9

	Heterosexual	MSM	Total
Subtype A	2	1	3
Subtype B	6	17	23
Subtype C	2	0	2
Subtype E	6	1	7
Total	16	19	35

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Fig. 10



4320

4361

4269

4081

4378

4589

4317

4494

4727

4309

4017

4441

4488

4480

4091

N1

P1

P2

N2

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DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that my residence, post office address and citizenship are as stated below next to my name: that I verily believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter claimed and for which a patent is sought in the application entitled:

METHOD FOR DETERMINING HIV-1 SUBTYPE

which application is:

☐ the attached application

(for original application)

☒ PCT International Application No. PCT/JF00/03896

filed on June 15, 2000

☐ Application No. _____

filed _____

, and amended on _____

(for declaration not accompanying application)

that I have reviewed and understand the contents of the specification of the above-identified application, including the claims, as amended by any amendment referred to above; that I acknowledge my duty to disclose information of which I am aware which is material to the patentability of this application under 37 C.F.R. 1.56, that I hereby claim priority benefits under Title 35, United States Code §119, §172 or §365 of any provisional application or foreign application(s) for patent or inventor's certificate listed below and have also identified on said list any foreign application for patent or inventor's certificate on this invention having a filing date before that of any foreign application on which priority is claimed:

Application Number

1999-167736

2000-23581

Country

Japan

Japan

Filing Date

15/06/1999

01/02/2000

Priority Claimed

Yes

No

☒

☐

I hereby claim the benefit of Title 35, United States Code §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in a listed prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge my duty to disclose any information material to the patentability of this application under 37 C.F.R. 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

Application No.

Filing Date

Status

I hereby appoint John H. Mion, Reg. No. 18,872; Thomas J. Macpeak, Reg. No. 19,292; Robert J. Seas, Jr., Reg. No. 21,092; Darryl Mexic, Reg. No. 23,063; Robert V. Sloan, Reg. No. 22,775; Peter D. Olexy, Reg. No. 24,513; J. Frank Osha, Reg. No. 24,625; Louis Gubinsky, Reg. No. 24,835; Neil B. Siegel, Reg. No. 25,200; David J. Cushing, Reg. No. 28,703; John R. Inge, Reg. No. 26,916; Joseph J. Ruch, Jr., Reg. No. 26,577; Sheldon I. Landsman, Reg. No. 25,430; Richard C. Turner, Reg. No. 29,710; Howard L. Bernstein, Reg. No. 25,665; Alan J. Kasper, Reg. No. 25,426; Kenneth J. Burchfiel, Reg. No. 31,333; Gordon Kit, Reg. No. 30,764; Susan J. Mack, Reg. No. 30,951; Frank L. Bernstein, Reg. No. 31,484; Mark Boland, Reg. No. 32,197; William H. Mandir, Reg. No. 32,156; Brian W. Hannon, Reg. No. 32,778; Abraham J. Rosner, Reg. No. 33,276; Bruce E. Kramer, Reg. No. 33,725; Paul F. Neils, Reg. No. 33,102; Brett S. Sylvester, Reg. No. 32,765; Robert M. Masters, Reg. No. 35,603; George F. Lehnigk, Reg. No. 36,359; John T. Callahan, Reg. No. 32,607; Steven M. Gruskin, Reg. No. 36,818; Peter A. McKenna, Reg. No. 38,551 and Edward F. Kennehan, Reg. No. 28,962, my attorneys to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith, and request that all correspondence about the application be addressed to SUGHRUE, MION, ZINN, MACPEAK & SEAS, PLLC, 2100 Pennsylvania Avenue, N.W., Washington, D.C. 20037-3213.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Date November 29, 2001

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State/Country

Signature

First Name

Middle Initial

Last Name

Post Office Address: 1-33-13, Miharadai, Nerima-ku, Tokyo 177-0031, Japan

Citizenship Japan

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SEQUENCE LISTING

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<110> KEIO UNIVERSITY

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10009897.1.21401

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